Lecture 4

Absorption Part 1 – theoretical basis of absorption, absorption by individual particles, bulk absorption

Collin Roesler

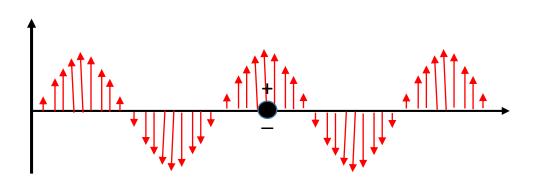
20 July 2021

Lecture Overview

- Overview of the electromagnetic spectrum
- Absorption on molecular scales
- Major absorbers in the ocean
- Lecture 5: Measuring absorption in the ocean
- Lab 2: Absorption by solutions
- Lab 3: Absorption by particles

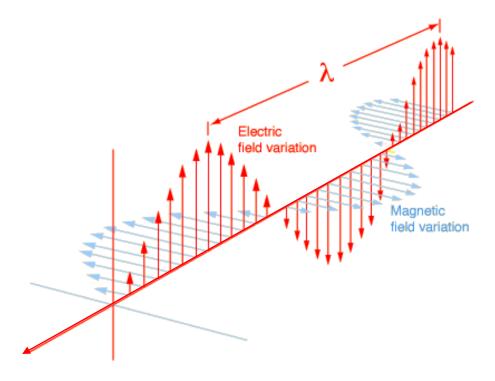
Electromagnetic Radiation

 Charged particles (dipoles) create electric fields E (oscillation between +,-)



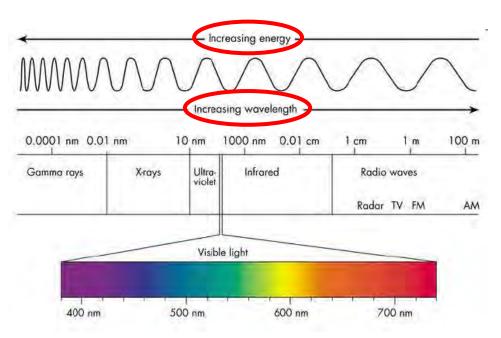
Electromagnetic Radiation

- Charged particles, dipoles, create electric fields E (oscillation between +,-)
- When a charged particle moves, it creates a magnetic field, B
- The electromagnetic field oscillates as the energy propagates **ExB** (right hand rule)



Electromagnetic Radiation

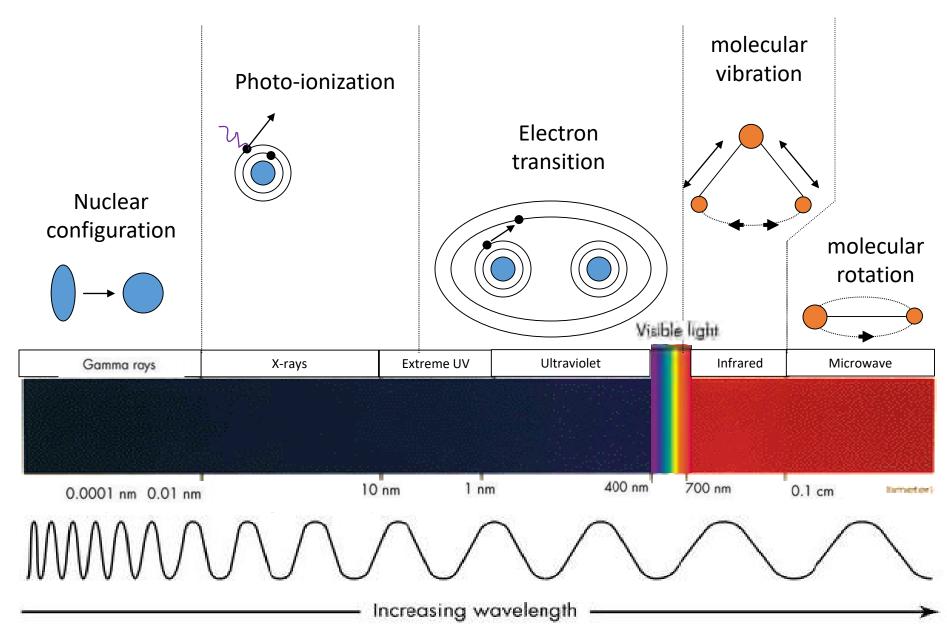
- Charged particles, dipoles, create electric fields E (oscillation between +,-)
- When a charged particle moves, it creates a magnetic field, **B**
- The electromagnetic field oscillates as the energy propagates **ExB** (right hand rule)
- the range of oscillation frequencies is described by the Electro-Magnetic (EM) spectrum



What is absorption?

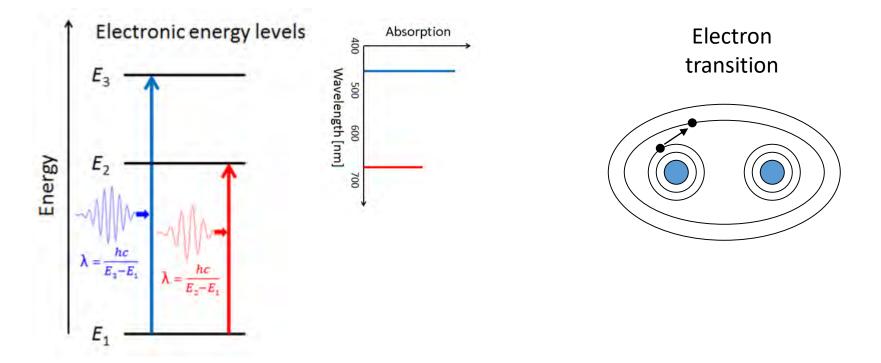
- Since electromagnetic radiation is energy propagation, when molecules absorb radiation, they absorb *energy*
- The energy associated with each part of the spectrum is given by $E = hc/\lambda$
 - h Planck's constant
 - c speed of light in a vacuum
 - λ wavelength
- What happens to the molecule depends upon the amount of energy, and therefore the wavelength

Interactions between energy and matter



Quantized electronic states

- Amount of energy required to move an electron to a higher orbital (electronic state transition) is *quantized*
- A molecule absorbs radiation **only** of this specific quantized energy
- The quantized energy determines the absorption peak wavelength
- This example shows a molecule with two higher energy levels (E2 and E3) and thus two absorption peaks

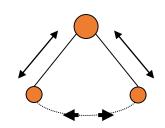


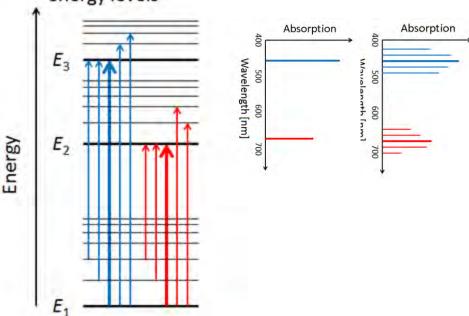
Quantized vibrational states

- Each orbital is associated with a series of higher excited states, associated with vibrational energy, which are also quantized
- These determine the wavelengths of the absorption side peaks that have higher or lower energy, but have a lower probability for absorption

Electronic + vibrational energy levels

molecular vibration

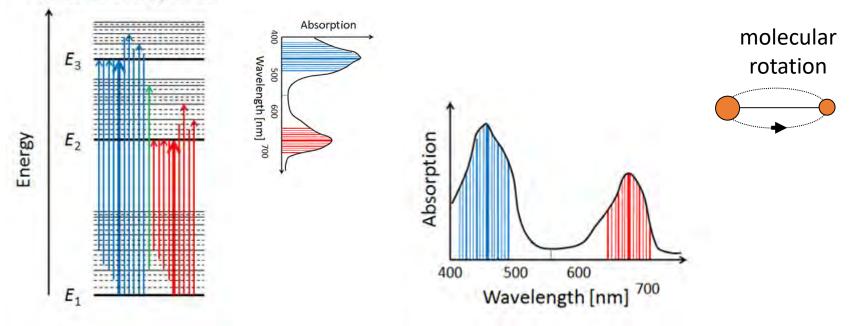




Quantized rotational states

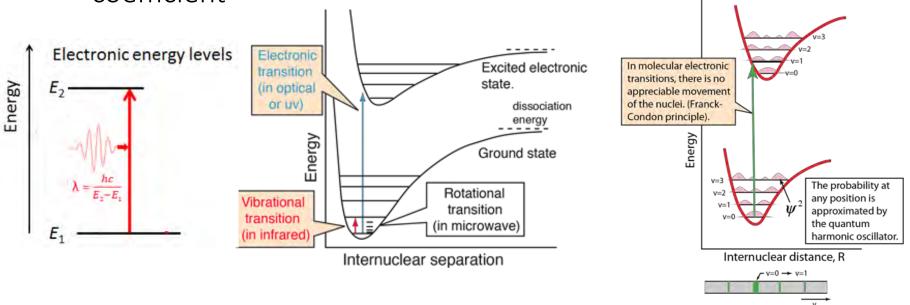
- Each vibrational state is associated with a series of higher rotational states, which are also quantized
- These determine the wavelengths of the absorption that smooth the absorption peaks

Electronic + vibrational + rotational energy levels

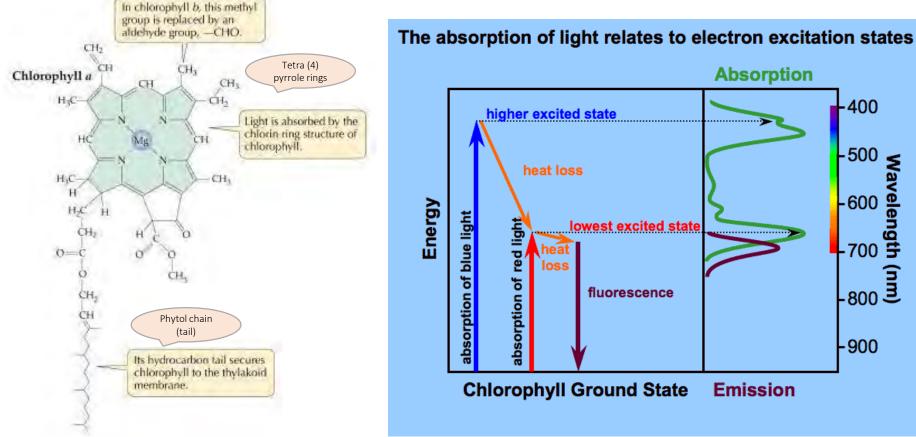


Quantized states

- Each level is really a distribution as a function of internuclear separation
- With probabilities (likelihoods of occurrence)
- Higher probability of transition → higher absorption coefficient

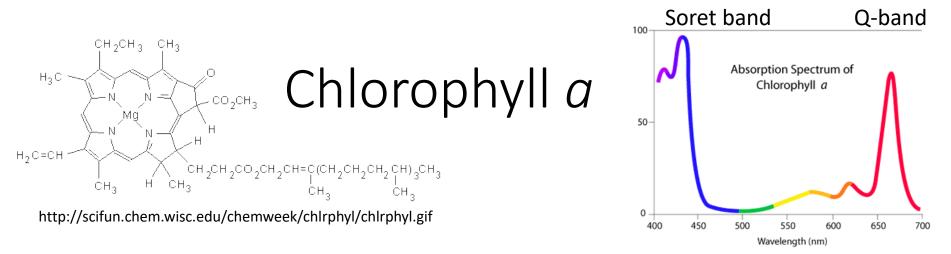


Chlorophyll *a* has two electronic states associated with the energy equivalent of blue (443 nm) and red (676 nm) wavelengths

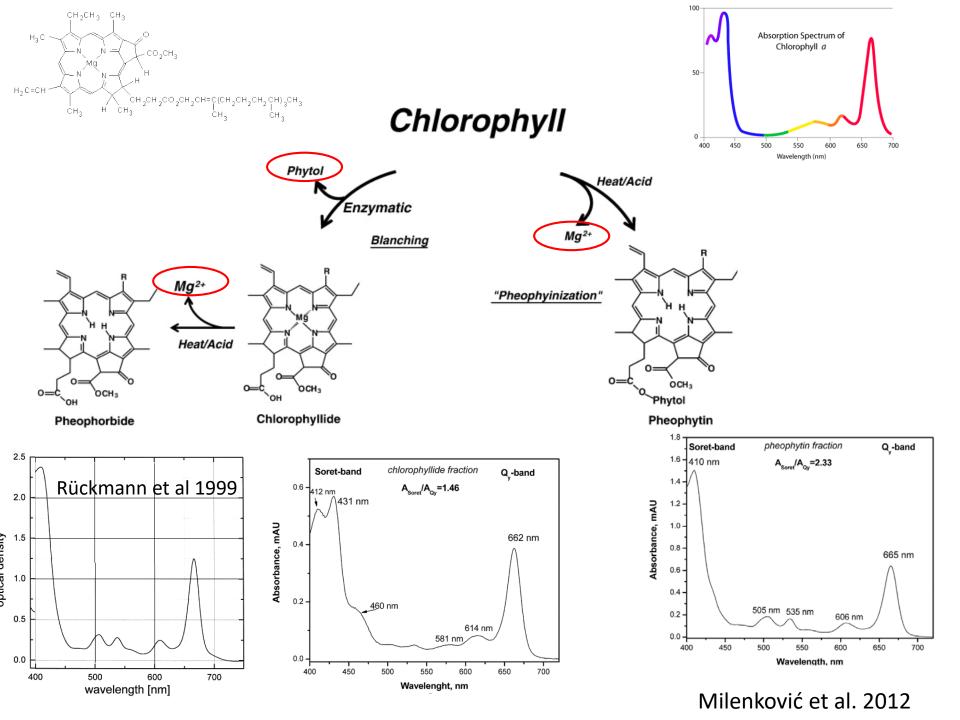


http://www.mie.utoronto.ca/labs

http://plantphys.info/plant_physiology/light.shtml



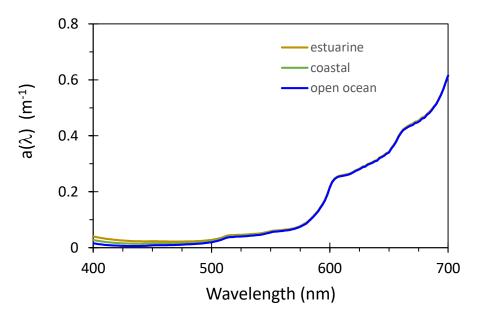
http://www.photochembgsu.com/assets/images/graph2.gif



What are the major absorbers in the ocean?

Example of absorption spectra for three environments

- What do they have in common?
 - All have strong red absorption
- How do they differ?
 - Variable blue absorption



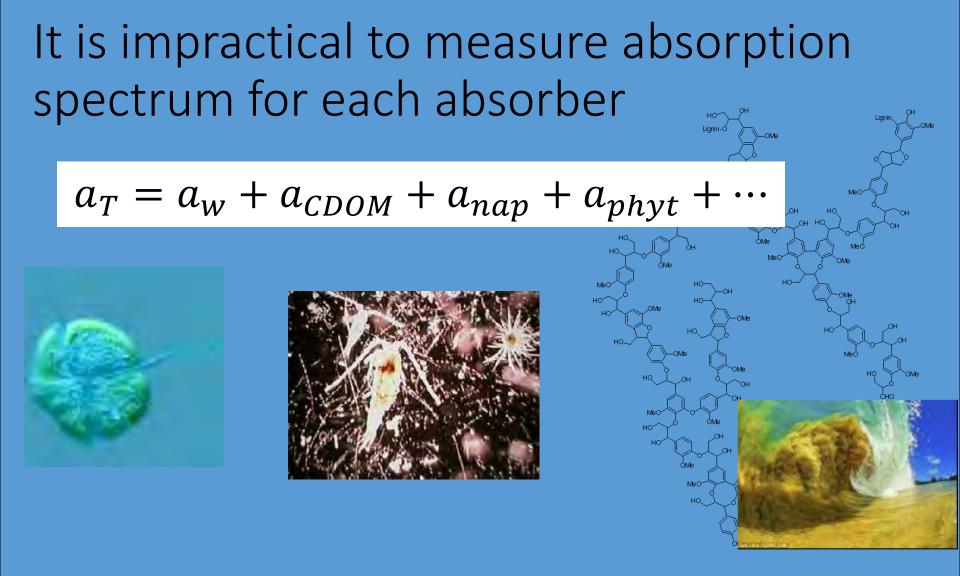
Absorption is a conservative property

• Total absorption = sum of individual absorbing constituents

$$a_{total} = a_{water} + \sum a_{dissolved} + \sum a_{particles}$$

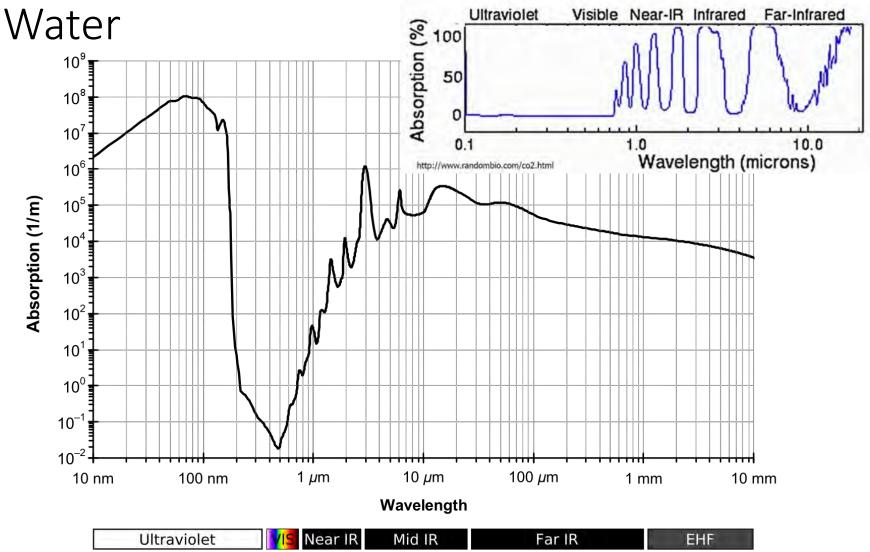
 Absorption is proportional to the concentration (Beer's Law)

$$a_{chl}(m^{-1}) = [chl](\frac{mg}{m^3}) \times a_{chl}^*(\frac{m^2}{mg})$$

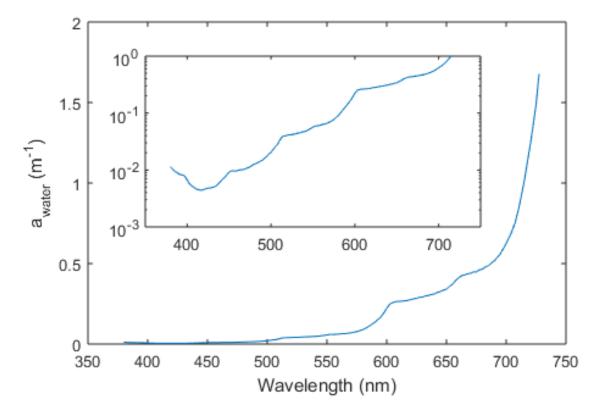


Group components by their common absorption properties (an our inability to separate them operationally)

Absorbing Components:



By Kebes at English Wikipedia, CC BY-SA 3.0, https://commons.wikimedia.org/w/index.php?curid=23793083



R. M. Pope and E. S. Fry 1997 Integrating cavity absorption meter

Nice (but dated) compendium at http://omlc.org/spectra/water/abs/index.html

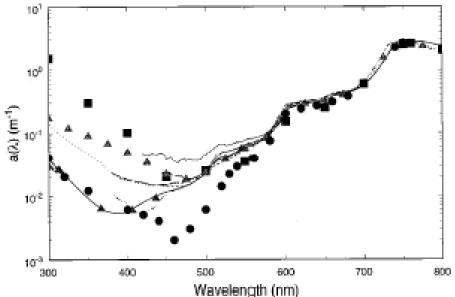
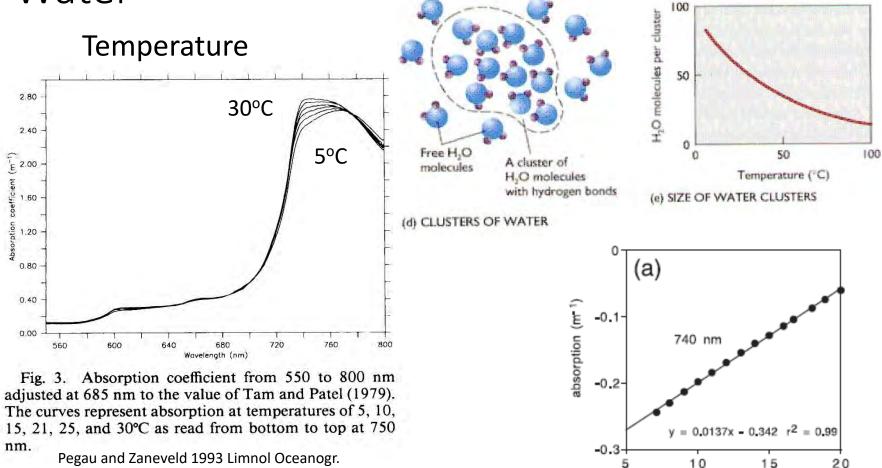


Fig. 1. Absorption coefficient of pure water as measured or compiled by several investigators.^{1,2,11,18,19,21,26–33} The discrepancy in the estimated absorption coefficients is largest at short wavelengths where absorption by organic contaminants is significant. At wavelengths longer than 550 nm the standard deviation of the estimates is between 5 and 10% of the mean value.

W. Scott Pegau, Deric Gray, and J. Ronald V. Zaneveld Absorption and attenuation of visible and near-infrared light in water: dependence on

temperature and salinity 20 August 1997 / Vol. 36, No. 24 / APPLIED OPTICS

variations are methodological



natural variations

Sullivan et al. 2006 Appl Opt

temperature (°C)

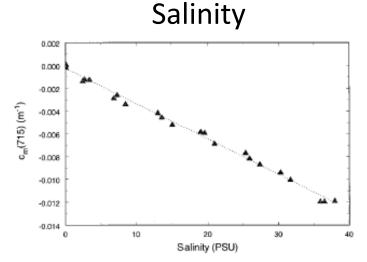
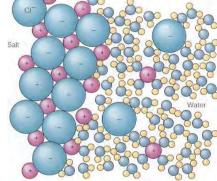
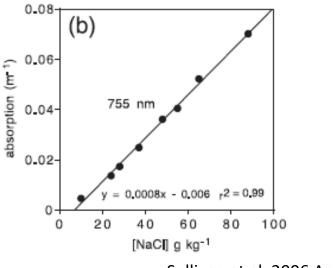


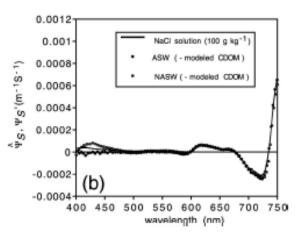
Fig. 6. Attenuation coefficient at 715 nm as a function of salinity. This figure illustrates the linear dependence of the attenuation coefficient on salinity. Pegau etal. 1997 Appl.Opt.



natural variations

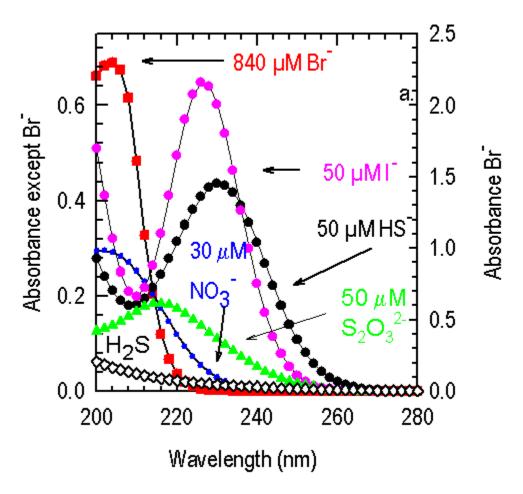


Sullivan et al. 2006 Appl Opt



Absorbing Components: Dissolved inorganic matter

- Basic for UV detection of nitrate, ISUS
- Johnson, K. S. and L. J. Coletti. 2002



http://www.mbari.org/chemsensor/ISUShome.htm

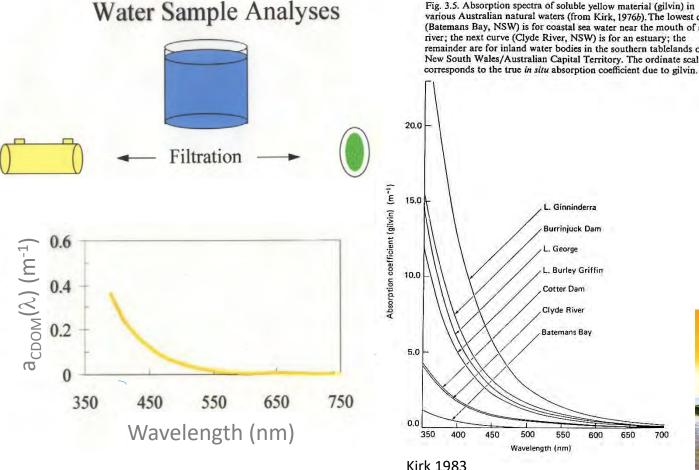


Fig. 3.5. Absorption spectra of soluble yellow material (gilvin) in various Australian natural waters (from Kirk, 1976b). The lowest curve (Batemans Bay, NSW) is for coastal sea water near the mouth of a river; the next curve (Clyde River, NSW) is for an estuary; the remainder are for inland water bodies in the southern tablelands of New South Wales/Australian Capital Territory. The ordinate scale

650

700



Dierssen et al. 2006



http://clarklittlephotography.com

Fig. 3.5. Absorption spectra of soluble yellow material (gilvin) in $a_{CDOM}(\lambda) = a_{CDOM}(\lambda_{ref}) \exp(-S_{CDOM}(\lambda - \lambda_{ref}))$ various Australian natural waters (from Kirk, 1976b). The lowest curve (Batemans Bay, NSW) is for coastal sea water near the mouth of a river; the next curve (Clyde River, NSW) is for an estuary; the remainder are for inland water bodies in the southern tablelands of New South Wales/Australian Capital Territory. The ordinate scale corresponds to the true in situ absorption coefficient due to gilvin. 20.0 ABSORPTION) LOOP CURRENT HUMIC Absorption coefficient (gilvin) {m⁻¹} 15.0 L. Ginninderra Burrinjuck Dam L. George OG (SPECIFIC MISSISSIPPI RIVER L. Burley Griffin 10.0 FULVIC LOOP CURRENT FULVIC Cotter Dam Clyde River Batemans Bay 5.0 400 200 300 500 600 700 WAVELENGTH (nm) 0.0 Fig. 1. Specific absorption curves vs. wavelength 350 400 450 500 550 600 650 700 Wavelength (nm) for marine humic acid and marine fulvic acid. Kirk 1983

Carder et al. 1989 L&O

$$a_{CDOM}(\lambda) = a_{CDOM}(\lambda_o) \exp(-S_{CDOM}(\lambda - \lambda_o))$$

Equatorial Pacific – filtrate pore size and spectral slope

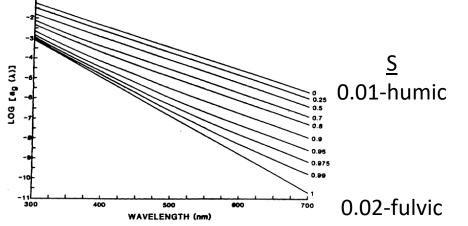


Fig. 3. Spectral variation of the absorption coefficient due to marine humus or Gelbstoff as a function of t^{\dagger} fulvic acid fraction of Gelbstoff for $a_{f}^{\bullet} = 0.00732 \text{ m}^{2} \text{ g}^{-1}$, $a_{h}^{\bullet} = 0.131 \text{ m}^{2} \text{ g}^{-1}$, $B_{f} = 0.0186 \text{ nm}^{-1}$, and $B = 0.0110 \text{ nm}^{-1}$. The fulvic acid fraction is shown beside each curve.

Carder et al. 1989 L&O

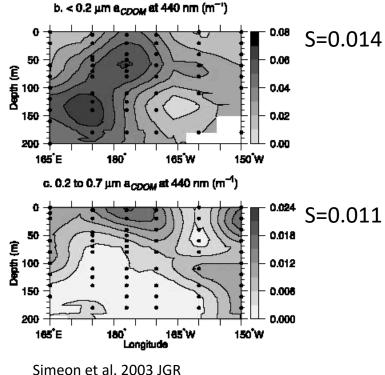
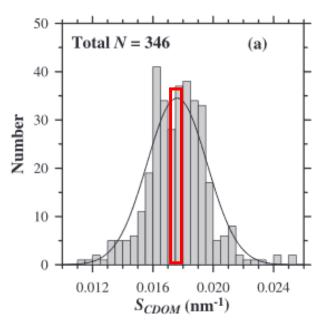


Table 1. Ranges for the exponential coefficient, $C2_{xx}$ for gelbstoff and detritus for Eq. 6. Where coefficients were not listed, values were approximated from published spectra using an exponential model.

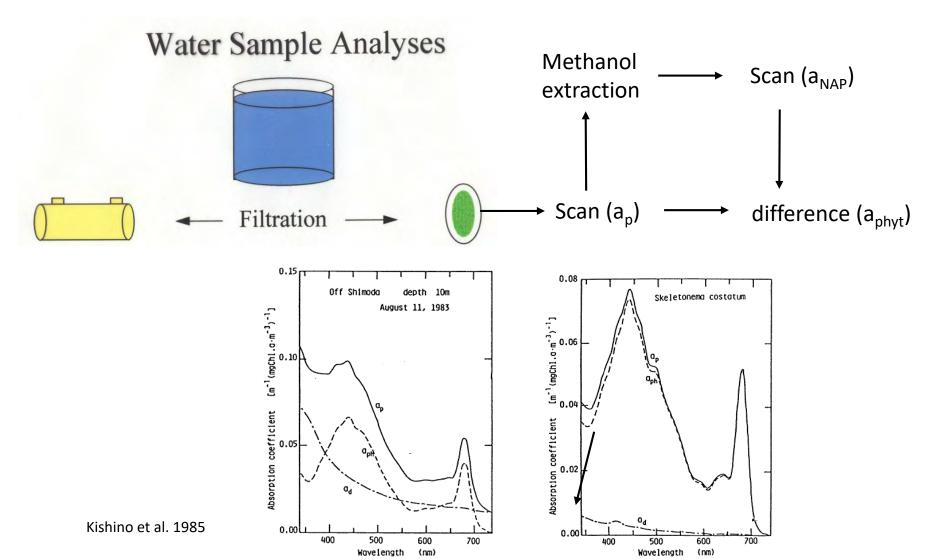
Reference	Site	Avg C2, (nm-i)
Gelbstoff		
Kalle 1966	Baltic, North Sea	0.018
Jerlov 1968		0.015
Kirk 1976	Lakes, coast	0.015
Lundgren 1976	Baltic	0.014
Kopelevich and Burenkov 1977	Indo-Pacific	0.017
Bricaud et al. 1981	Baltic	0.018
	Mauritania	0.015
	Gulf of Guinea	0.014
	Mediterranean	0.014
Okami et al. 1982	East Pacific	0.017
Kishino et al. 1984	Lake Kizaki	0.016
	Nabeta Bay	0.015
	East Pacific	0.014
Carder and Steward 1985	Gulf of Mexico	0.014
Davies-Colley and Vant 1987	Lakes	0.019
Maske and Haardt 1987	KICI Harbor	0.016
Published mean ± SD		0.016 ± 0.002
This study mean ± SD	San Juan Islands	0.017 ± 0.003
Caroor et al. 1707	Marine numic acid	0.011
Detritus	Marine fulvic acid	0.018
Kishino et al. 1986	NW Pacific Ocean	0.006
Maske and Haardt 1987	Kiel Harbor	0.014
Iturriaga and Siegel 1988	Sargasso Sea	0.011
Cleveland and Perry in prep.	Sargasso Sea	0.013
Morrow et al. 1989	Sargasso Sea	0.009
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Babin et al. 2003 (European coastal waters)

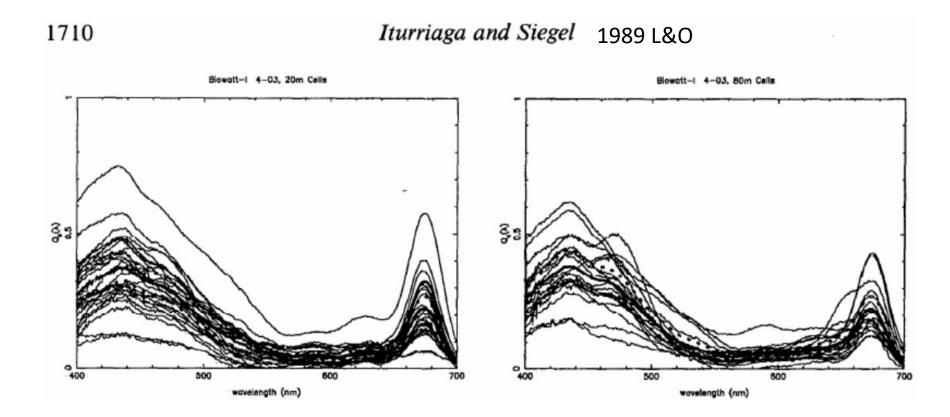
Roesler et al. 1989 (global synthesis)

Absorbing Components: Particles

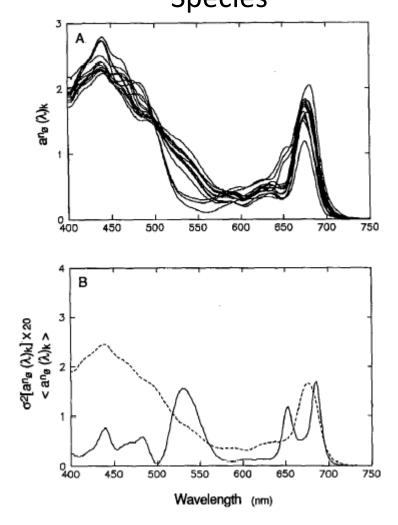


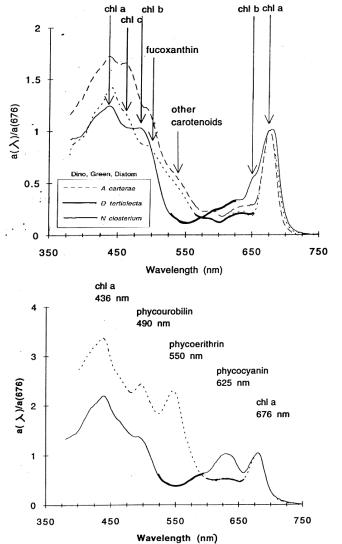
Absorbing Components: Phytoplankton

Individual cells, microphotometry



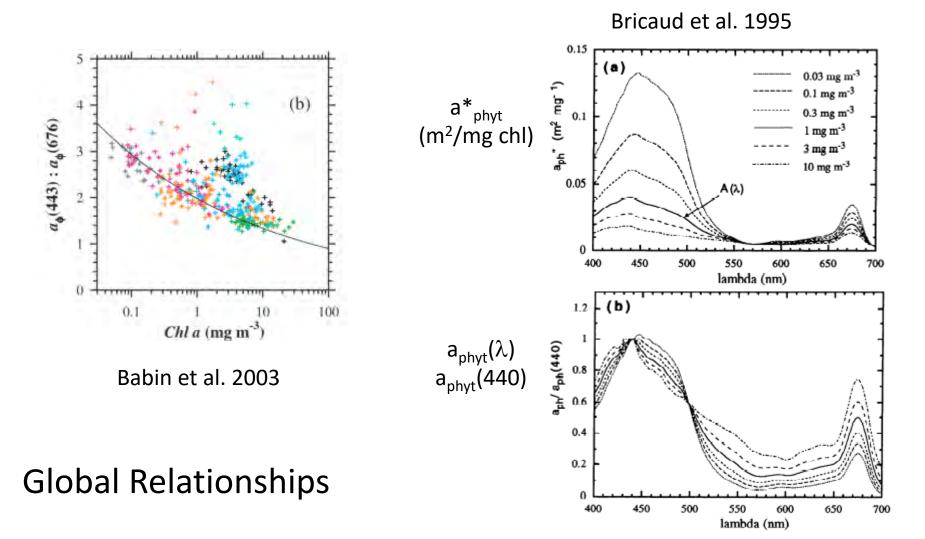
Absorbing Components: Phytoplankton Species



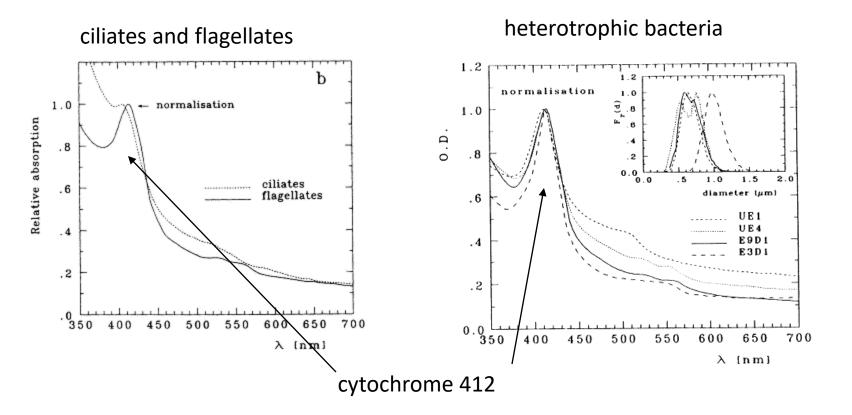


Roesler et al. 1989 L&O

Absorbing Components: Phytoplankton



Absorbing Components: other protists

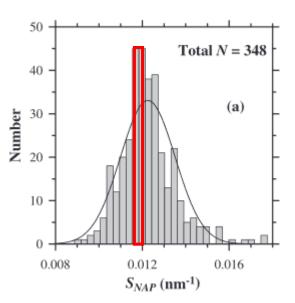


Morel and Ahn 1990 JMR

Absorbing Components: Non-Algal Particles

Table 1. Ranges for the exponential coefficient, $C2_x$, for gelbstoff and detritus for Eq. 6. Where coefficients were not listed, values were approximated from published spectra using an exponential model.

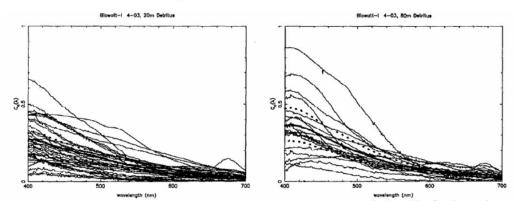
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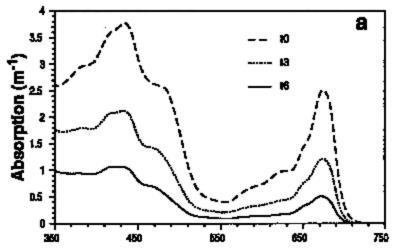
Babin et al. 2003 (European coastal waters)

Roesler et al. 1989 (global synthesis)

Absorbing Components: Non-algal particles \rightarrow what are they?



Nelson & Robertson: Detrital spectral absorption 1993] JMR



Iturriaga and Siegel 1989 L&O

Photobleaching natural light levels

Absorbing Components: inorganic particles

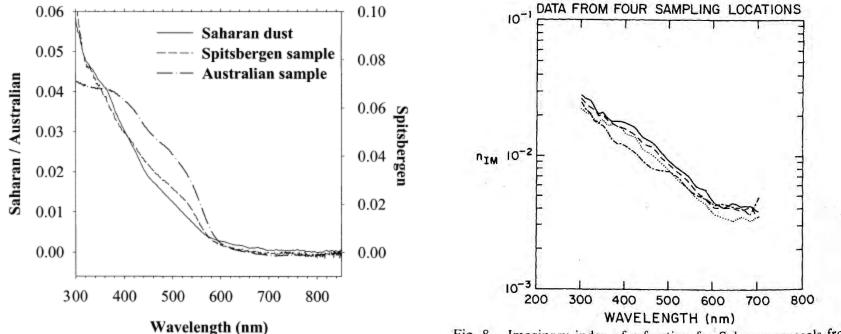


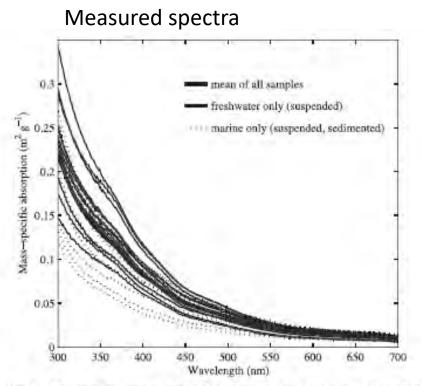
Fig. 5. Absorbance spectra of natural assemblages of mineral particles from three different environments.

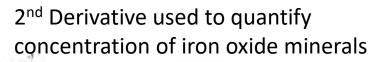
Babin and Stramski 2003

Fig. 8. Imaginary index of refraction for Saharan aerosols from each of the collection locations. The solid line represents the Tenerife sample; the dashed line, the *Meteor* sample; the dotted line, the Barbados sample; and the dashed-dotted line, the Sal Island sample.

Patterson et al. 1977 JGR

Absorbing Components: inorganic particles





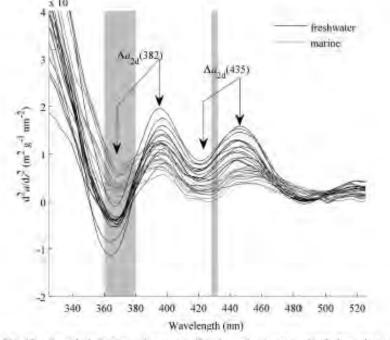
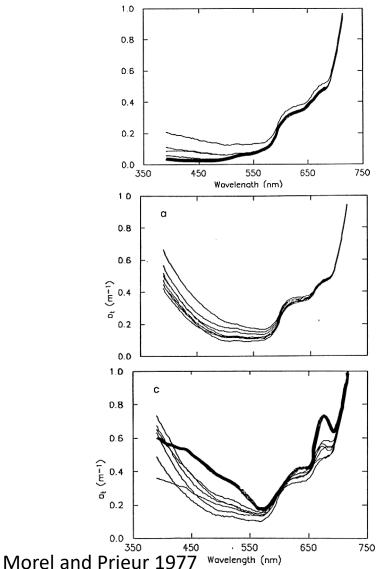


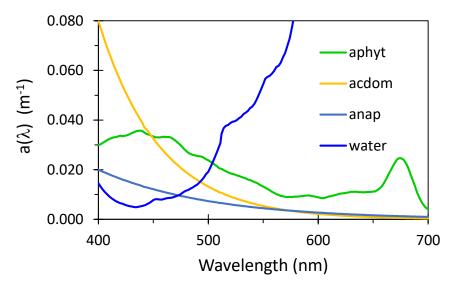
Fig. 3. Mass-specific absorption spectra of all samples analyzed here (n = 25). Heavy black line shows the mean, this solid lines show samples from freshwater sites on the Atchafalaya and Mississippi Rivers, and dashed lines show samples from marine sites at Freshwater Bayou and the Atchafalaya River delta. River samples are suspended particulates only, marine samples include both sediments and suspended particulates.

Fig. 10. Second derivatives of main-specific absorption spectra. Black lines denote freshwater samples, and gray lines denote marine samples. Bracketed arrows labeled $\Delta a_{2d}(382)$ and $\Delta a_{2d}(435)$ show locations of second-derivative maxima and minima used to compute iron absorption peak heights plotted in Fig. 11. Light-gray vertical bars highlight approximate marges for electronic transition bands of various iron oxide minerals (Sherman and Waite 1985).

Estapa et al. 2012

To model the impacts of absorbing constituents \rightarrow add them up





Which component dominates?

- blue waters
- green waters

phytoplankton (V-type) inorganic particles (U-type)

More on absorption

- Today
 - Lecture 5: Measuring absorption (Collin)
 - Lab 2 CDOM absorption methods
- Wednesday
 - Lab 3 Particulate absorption methods