

Fluorescence class at DMC-- 22 October 2012

HANDS-ON:

Meet in Perry lab and head to the dock:

- 1) water sampling from the dock
 - 2 students sample three replicates (~ 130 mL)
 - one student – sample three volumes (~ 60, 130, 280 mL)
 - remaining students – one sample (~ 130 mL)
- 2) BB2F on the dock, during water sampling
- 3) net tow to collect phytoplankton

Perry lab:

- 1) Filter water samples and put in acetone (Ivona Cetinic will sonicate).
- 2) Tour of Perry lab – Turner Designs fluorometer; lab safety
- 3) spinach extract – visual of chlorophyll fluorescence

Mayer Lab:

At end of class – if time – back to the Perry lab to read the chlorophyll extracts.

LECTURE/DISCUSSION:

- 1) Who in the class has used chlorophyll fluorescence? When? Why? How?
- 2) Big picture
 - what do we really want to know about phytoplankton?
 - o (biomass, number of phytoplankton, size distribution, species, ?)?
 - why the interest in chlorophyll fluorescence?
 - o you might want it to find chlorophyll concentration
 - o is chlorophyll concentration a good proxy for phytoplankton?
 - o proxy for phytoplankton what?
 - 'biomass', but requires chlorophyll to element ratio (Chl/C)
 - does not give cell number
 - phytoplankton size or size distribution?
 - maybe, with size fractionations of chlorophyll (Nitex)
 - maybe mean size with variance/mean ratio (Nathan)
 - what does fluorescence actually provide?
 - do coupled measurements get us closer? (other optical measurements – bbp)
- 3) Overview – different ways to think about phytoplankton (molecule, membrane, cell)
- 4) Fluorescence in general and chlorophyll fluorescence in particular; relationship between absorption and fluorescence

- 5) BRIEF overview and history of measurement of chlorophyll – or different aspects of chlorophyll (place fluorescence in context and reference for ‘calibration’)
 - *in vivo*: by eye (color of the water) and by smell
 - *in vivo*: by eye (Munsell color chart)
 - *in vitro*: spectrophotometry of extracts
 - *in vitro*: fluorescence of extracts
 - *in vitro*: HPLC of extracts
 - *in vivo*: chlorophyll fluorescence profiles by pump, then *in situ* fluorometry
 - *in vivo*: epifluorescence
 - *in vitro*: spectrophotometry of filter pads (a_676)
 - *in vivo*: spectrophotometry with ac9/acs (a_676), profiles and underway
 - *in vivo*: flow cytometry on bench and *in situ*
 - remote sensing: ocean color remote sensing reflectance
 - remote sensing: ocean color remote sensing fluorescence line height
 - *in vivo*: pump and probe fluorescence for physiology (profiles and bench top)
- 6) general principles of fluorometric measurement - excitation/emission, light source, filters, detectors, geometry, temporal resolution.
 - bench top and *in situ* manufacturers
 - choice of wavelength (absorption spectra, excitation/emission spectra)
 - calibration for fluorometers – *in vitro* (extracts) and *in vivo* (of living cells); what is calibrated?
 - drift – electronics, filter degradation
 - temperature effects – electronics, fluorescence quenching
- 7) interpretation of data, and challenges therein, and potential workarounds
 - fluorescence quenching due to sun and innate diel rhythms
 - pigment packaging – cell size and photoadaptation
 - variable ratio of photosynthetic pigments
 - nutrient limitation
- 8) synthesis

Assigned readings

Basic readings on fluorescence and fluorometers in general, and chlorophyll fluorescence in particular:

Turner Designs– the Basics.pdf

Turner Designs website - practical info.docx

Primer on fluorescence.pdf (some redundancy with Turner Designs – the Basics)

Suggett et al. 2010 – table of contents.pdf (Chapters 3 & 7 are from Suggett et al. – available as an e-book from UMaine library)

Historical use of chlorophyll fluorescence – the classic paper:

Lorenzen 1966.pdf

The method you will use to measure chlorophyll concentration:

JGOFS_chl_method.pdf

Additional reading, if you are interested, mostly on ‘variable fluorescence’:

Chapter 3 Huot Babin overview.pdf

Chapter 7 Richardson taxonomic discrimination.pdf