

Oxygen and the photodissolution of shallow coastal suspended sediments and phytoplankton detritus

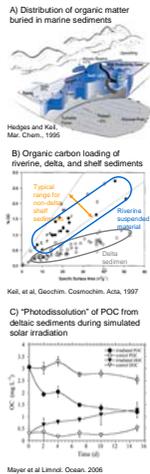
Margaret L. Estapa, Lawrence M. Mayer, and Linda L. Schick

(margaret.estapa@maine.edu)

University of Maine, School of Marine Sciences

Background

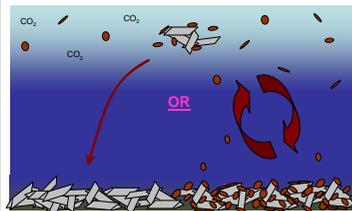
- River delta sediments account for ~44% of organic carbon (OC) buried in marine sediments, globally. (Hedges and Keil 1995). (A, upper right)
- However, deltaic sediments have lower surface-area normalized OC loadings than suspended riverine and non-deltaic shelf sediments (Keil *et al* 1997). (B, middle right)
- “Photodissolution” of POC from resuspended deltaic sediments is one hypothesis consistent with their relatively lower OC loadings (Mayer *et al* 2006). (C, lower right)
- 25-40% of POC is dissolved during laboratory irradiations. Typically, 5% is not detected as DOC.**



Questions

Is photodissolution reversible, once particles settle out of the surface water (right side of schematic, below)?

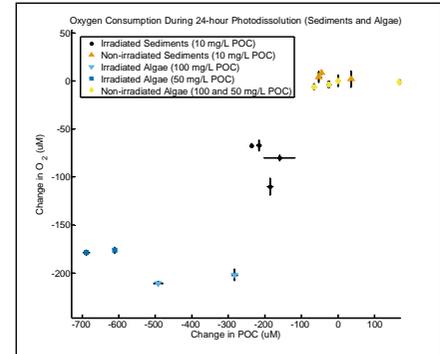
Is photodissolved POC *photooxidized*, as is dissolved organic carbon (left side of schematic)?



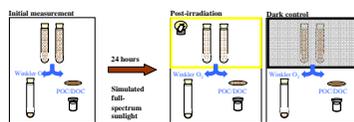
What are impacts on O₂ and nitrogen forms during photodissolution?

Results and Discussion

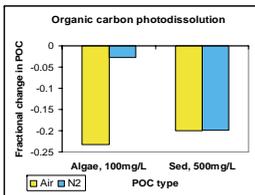
- Sediments at reasonable field concentrations resulted in strong depletions of O₂. For suspensions with the highest initial POC concentrations (algae, 100 mg/L), O₂ levels after 24 h of irradiation were below detection.
- For both algal membrane and Atchafalaya Bay bottom sediments, the photodissolution of POC consumed O₂ at a molar ratio (C:O₂) of about 3:1. This is consistent with the remineralization of 5% of total organic carbon not accounted for after irradiation.



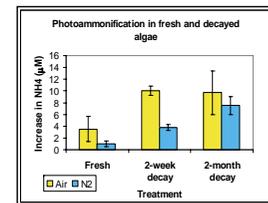
Study Area and Experimental Design



- Suspensions of Atchafalaya bottom sediments and freeze-dried algae (*Tetraselmis* spp., membrane fraction) made up in carbon-free artificial seawater.
- Replicate suspensions were irradiated in a solar simulator or kept in the dark as non-irradiated controls, all at room temperature.
- Suspensions were analyzed for POC, DOC, and dissolved O₂ before and after irradiation.
- To test for effects of microbial degradation followed by irradiation, additional *Tetraselmis* membrane samples were allowed to decay for periods of 2 weeks and 2 months prior to irradiation and analysis as described above. Additional analyses for DON, PN, total acid- and enzymatically-hydrolyzable amino acids (THAA and EHAA), NO₃⁻, and NH₄⁺ were carried out on these samples.

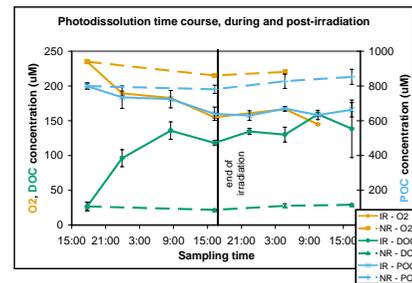
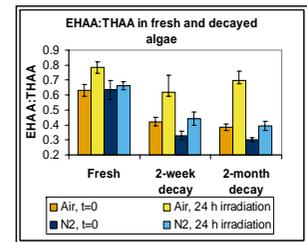


- Algal membrane photodissolution was strongly inhibited in N₂-saturated suspension, but reaction extents were nearly equal for air- and N₂-saturated sediment suspensions.



- Photoammonification occurred during irradiation of fresh and decayed algal membranes, making up 12-18% of total N photodissolution. Production was higher for decayed membranes and underoxic conditions.

- The EHAA:THAA fraction of THAA decreased during pre-irradiation algal decay. Irradiation then increased access of enzymes to THAA, suggesting that the initial drop was due to protein “encapsulation” by protective material or cross-linking that is especially sensitive to irradiation under O₂.



- POC, DOC, and O₂ monitored for further 24 h after the end of a 24-h irradiation of sediments exhibited no significant additional dissolution, oxygen consumption, or readsorption, consistent with irreversibility of the photodissolution process.

Conclusions

- O₂ is involved, when present, in the photodissolution of POC from suspended sediments.
- O₂ is likely required for the photodissolution of fresh algal photodetritus.
- The C:O₂ molar loss ratio is about 3:1, consistent with photo-oxidation of the fraction of photodissolved POC not detected as DOC.
- Irradiation may accelerate N cycling, via enhanced hydrolysis and photoammonification.

References

Hedges JI and RG Keil, 1995. *Mar. Chem.* **49**: 81-115.
Keil RG, et al, 1997. *Geochim. Cosmochim. Acta* **61**(7): 1507-1511.
Mayer LM, et al, 2006. *Limnol. Oceanogr.* **51**(2): 1064-1071.

Acknowledgements

We'd like to thank Kathy Hardy, who performed the dissolved nitrogen analyses for this work.

This work is supported by the NSF Chemical Oceanography program

