Absorption, Part 1

- Context (in this class, and broader disciplinary context)
- How does light energy interact with matter in the ocean?
- Overview of absorbing constituents in the ocean

Class context: Week 1 roadmap



Disciplinary context

- Undergraduate physical chemistry connections (e.g., spectroscopy)
- Language to help you read about these concepts outside the field of oceanography and/or talk about them with non-oceanographers
- Foundation to support your future work in new directions that none of us have considered, yet!

Absorption, Part 1

- Context (in this class, and broader disciplinary context)
- How does light energy interact with matter in the ocean?
 - Electromagnetic spectrum
 - Quantized absorption energies
 - Fate of energy after absorption
- Overview of absorbing constituents in the ocean

Electromagnetic (E-M) energy spectrum







https://www.noaa.gov/jetstream/satellites/absorb

Examine the **spectral shape of the relative absorption** of incoming sunlight by Earth's atmosphere.

What notable features do you see?

Bands in NIR Distinct Large absorption/Scattury in UV S Decreasing intensity of IR bands with decreasing 2



Examine the **spectral shape of the relative absorption** of incoming sunlight by Earth's atmosphere.

What notable features do you see?

- Light is absorbed in discrete wavelength bands of varying width
 - Absorption occurs only at specific, quantized energies
- Light is strongly absorbed at ultraviolet wavelengths
 - UV light excites electrons (eg, O₃ bonds)
 - Shorter wavelengths undergo more Rayleigh scattering



Molecular Orbitals for Water Vapor

Electronic transitions

- Red and blue blobs are molecular orbitals, the "probability regions" where the electrons are found around the molecule
- Electrons can *only* occupy these regions
- Going from bottom to top, the orbitals are arranged from lowest to highest energy (theoretical values on the right)
- For an electron to change shape between orbitals, it must absorb or release a photon with energy equal to the *difference* between those orbitals

https://commons.wikimedia.org/wiki/File:Molecular_Orbitals_for_Water.png



Molecular Orbitals for Water

Electronic transitions

- In its "ground" or lowest-energy state, water's 10 electrons pair up in the bottom 5 orbitals
- If a water vapor molecule absorbs a photon with λ = 166.5 nm then an electron can be *excited* from the 5th to the 6th orbital in the diagram





Electronic transitions

- Most of the time we don't know the actual orbital shapes, instead we measure the energies that separate the states
- And often we don't bother showing states that don't interact with the part of the EM spectrum we're interested in





Electronic transitions

Vibrational states of a molecule



Simulation shows total vibrational and rotational states



http://mw.concord.org/nextgen/#interactives/chemistry/inter molecular-attractions/hydrogen-bonds

Vibrational "modes" of a molecule

- Vibrational states include fundamental modes animated here, plus harmonics and higher order combinations
- The larger and more complex the molecule, the more possibilities there are...





Animations:

https://www.oceanopticsbook.info/view/absorption/physics-ofabsorption



Electronic + vibrational + rotational? transitions



Factors that may modify absorbed energies

- State of matter (gas/liquid/solid)
- Internuclear distance
 - Temperature
 - Salinity
 - pH
 - Solvent effects



What notable differences do you see?

Vapor is different - More variable Vapor peaks are Shifted

https://commons.wikimedia.org/wiki/File:Water infrared absorption coefficient large.gif

State of matter (water)



https://commons.wikimedia.org/wiki/File:Water infrared absorption coefficient large.gif

What notable differences do you see?

 Liquid water and ice don't have the rotational structure. Molecules in liquids and solids can't rotate as easily.

•

The wavelengths of the vibrational transitions are not the same. Absorption energy depends on internuclear distance.

[Note: Narrow troughs in the middle of some vapor absorption bands shown here (e.g. 6.3 μm) probably arise because the absorption spectrum was derived from an atmospheric profile with a temperature inversion.]

Internuclear distance and transition probability

- Franck-Condon principle: Nuclei are effectively motionless during the time it takes for an electron to change state
- The more two states are "in phase", the higher the probability of an absorption (or fluorescence! more later) event
- It follows that anything affecting internuclear distance can alter the absorption energies.



https://chem.libretexts.org/Bookshelves/Physical and Theoretical Chemistry Textbook Maps /Physical Chemistry (LibreTexts)/13%3A Molecular Spectroscopy/13.07%3A The Franck-Condon Principle





Figure 8.6: Dependence of the water absorption coefficient on temperature and salinity according to Roettgers et al. (2014).

pH dependence of absorption

- pH dependence of carbonnormalized absorption by colored dissolved organic matter (CDOM) isolated from the macroalgae Sargassum
- Changes broadly attributable to loss of protons from phenolic and carboxylic groups
- ∆a_{pH} peak at 315 nm may be unique to brown algae?

Powers et al., 2020. https://www.frontiersin.org/articles/10.3389 /fmars.2020.588287/full



Solvent effects: Extracted vs. in vivo pigments



Figure 8.10, Mobley et al., *The Oceanic Optics Book*, 2022.

- 1. Pigment is "packaged" in the cell (more on this later)
- 2. Pigment absorption bands are shifted due to chemical environment in solvent, vs. in vivo.

Table 1. Absorption maxima, extinction coefficients (E) and wavelength shifts used for generating the "unpackaged" in vivo specific absorption coefficients for the major algal pigment types (PE = phycoerythrin from specific clones of Synechococcus sp.). The absorption maxima reported for the chlorophylls and carotenoids were determined in this study.

Pigment	λ_{\max} (solvent)	C (L g ⁻¹ cm ⁻¹)	λ shift
Chlorophyll a	662 nm (acetone) 432 nm	88.1512	>550 nm: +12 nm <550 nm: +8 nm
Chlorophyll b	646 nm (90% acetone) 458 nm	51.3612	>550 nm: +6 nm <550 nm: +12 nm
Chlorophyll c1+2	630 nm (90% acetone) 444 nm	42.612	>550 nm: +6 nm <550 nm: +16 nm
Fucoxanthin	450 nm (EtOH)	160.013	>400 nm: +40 nm
β , β -carotene	452 nm (EtOH)	262.014	>400 nm: +10 nm
WH7803 PE	547 nm (Pi buffer) ¹¹	25,515	no shift
WH8103 PE	492 nm (Pi buffer) ¹⁶	11.616	no shift

Table from Bidigare et al., 1990. doi:10.1117/12.21451

• Absorption: ~10⁻¹⁵ - 10⁻¹³ s



Fig. 7.9, Mobley et al., The Oceanic Optics Book, 2022.

- Absorption: ~10⁻¹⁵ 10⁻¹³ s
- Rayleigh scattering: ~10⁻¹³ s
 - Photon re-emission happens quickly, faster than vibration can occur
 - "Elastic scattering" = no change in wavelength (more tomorrow!)



Fig. 7.9, Mobley et al., The Oceanic Optics Book, 2022.

- Absorption: ~10⁻¹⁵ 10⁻¹³ s
- Rayleigh scattering: ~10⁻¹³ s
- Raman scattering: ~10⁻¹³ s
 - Also very fast before molecule can vibrate
 - "Inelastic" scattering
 - For water, characteristic shift in emitted photon of ~3400 cm⁻¹ (tens to hundreds of nm)



Fig. 7.9, Mobley et al., *The Oceanic Optics Book*, 2022. (addition of Raman scatter is not in the Mobley figure – errors are mine!)

- Absorption: ~10⁻¹⁵ 10⁻¹³ s
- Rayleigh scattering: ~10⁻¹³ s
- Raman scattering: ~10⁻¹³ s
- Fluorescence: ~10⁻⁹-10⁻⁷ s
 - Also "inelastic" scatter
 - Happens after vibrational relaxation – takes longer
 - Example: dissolved organic matter fluorescence. Also chlorophyll but biology complicates this example (more Friday)



Fig. 7.9 et al., Mobley, The Oceanic Optics Book, 2022.

- Absorption: ~10⁻¹⁵ 10⁻¹³ s
- Rayleigh scattering: ~10⁻¹³ s
- Raman scattering: ~10⁻¹³ s
- Fluorescence: ~10⁻⁹-10⁻⁷ s
- Phosphorescence: >>10⁻⁹ s
 - Electron spin flip (intersystem crossing) leads to a "forbidden transition" and much slower re-emission of light
 - Example = glow-in-the-dark Tshirts



Fig. 7.9, Mobley et al., The Oceanic Optics Book, 2022.

- Absorption: ~10⁻¹⁵ 10⁻¹³ s
- Rayleigh scattering: ~10⁻¹³ s
- Raman scattering: ~10⁻¹³ s
- Fluorescence: ~10⁻⁹-10⁻⁷ s
- Phosphorescence: >>10⁻⁹ s
- Photochemical reaction
 - No re-emitted light instead excited state molecule undergoes a chemical reaction
 - Examples: photosynthesis, photooxidation of CDOM



Fig. 7.9, Mobley et al., The Oceanic Optics Book, 2022.

Absorption, Part 1

- Context (in this class, and broader disciplinary context)
- How does light energy interact with matter in the ocean?
- Overview of absorbing constituents in the ocean
 - Water
 - Colored "dissolved" [[in?]organic?] matter (C"D"[O?]M)
 - Non-pigment particulate absorption (aka "non-algal particles", NAP)
 - Phytoplankton pigments

Water

- Spectral absorption coefficient of water, a_w(λ) [m⁻¹]
- Arises mainly from molecular vibrations (T, S dependence!) at visible-IR
- Difficult measurements to make...



Pope and Fry, 1997. Appl. Opt. 36(33): 8710-8723.



Figure 5.1 from Mobley et al., *The Oceanic Optics Book*, 2022. Data are from Pope and Fry, 1997.





Issues arising in measurements of pure

Fig. 11. Absorption coefficient, α , of pure water measured in this study along with the results from several other high-quality studies: Pope and Fry [8], the Rayleigh-scattering-corrected Quickenden and Irvin values [11], Cruz *et al.* [10], Lee *et al.* [30], and Morel *et al.* Extrapolation [17], and data at 10-nm intervals from Kröckel and Schmidt [14].

Both figures are from Mason, Cone, and Fry, 2016. 10.1364/AO.55.007163

Correcting for temperature and salinity effects

- Temperature dependence Ψ_T [m⁻¹ °C⁻¹]
- Salinity dependence (Ψ_{s}) [m⁻¹ S⁻¹]
- Recall the salinity dependence arises from salt ions modifying the water absorption – not intrinsic absorption by the salt ions!
- Dependences in the UV...?



Figure: Sullivan et al., 2006. Appl. Opt. 45(21): 5294-5309.

• What is "dissolved"?

What is "dissolved"?

- Smaller than filter size
- Fully "solvated" (all parts of the molecule interact with solvent)
- Gel assembly?

 terdence to stan suspended in still water
 → Always state your assumptions...

Figure: Libes, Introduction to Marine Biogeochemistry, 2009. Figure 22.1. ISBN 978-0-12-088530-5



- What is "dissolved"?
- What is "organic"?
 - Are all absorbing materials in the ocean (besides water) "organic"?

Optical nitrate sensor (ISUS, SUNA)

- Measurement principle: If concentration-specific UV absorption by NO₃⁻ is known, then measure absorption and calculate concentration (Beer's Law – more later).
- Sea salt interferences: bromide (Br⁻), nitrite (NO₂⁻), and bisulfide (HS⁻)
- Non sea-salt interference: CDOM
- Nitrite has a similar absorption maximum to nitrate but [NO₂⁻] is usually small (except where oxygen is low...)

Johnson and Coletti, 2002. *Deep-Sea Res. I*, 49: 1291-1305.



Fig. 1. Absorption spectra of bromide, bisulfide, nitrate and nitrite at concentrations typical of seawater. Each salt was dissolved in deionized water. Spectra were collected with a Hewlett-Packard HP 8452 spectrophotometer. Cell path length was 1 cm. Note that the left axis applies to bromide and the right axis to the other species.



Why is bromide absorption pressure-dependent? What other molecules could behave the same way?

Figures: Sakamoto et al., 2017. 10.1002/lom3.10209

2000

30

40

20

Nitrate (μ M)

10

0

- What is "dissolved"? *However you define it.*
- Are all absorbing materials in the ocean* (besides water) "organic"? No! Be aware of inorganic absorbers, in your spectral region of interest.



1 = No color, N and P absent (e.g., carbohydrates)
2 = Colored but N and P absent (e.g., carotenoids)
3 = Fluorescent but N and P absent (e.g., vanillic acid)
4 = Fluorescent and contains N (e.g., tyrosine)
5 = Fluorescent and contains both N and P (e.g., NADH)

Figure: Stedmon and Nelson, 2015, *Biogeochemistry of Marine Dissolved Organic Matter, 2nd ed.*

• OK, now we can talk about CDOM.

* In some seasonally-anoxic lakes also consider iron!

Generic CDOM absorption spectrum

Why is the spectrum smooth? Two hypotheses have been proposed:

- It is a superposition of many conjugated "pi" bonds with a range of energies (proposed by Shifrin, 1988)
- It arises from intramolecular charge transfer along a series of coupled, excited chromophores (Blough and Del Vecchio, 2004)





Figure 5.4. Mobley et al., 2022. The Oceanic Optics Book

Figure: Del Vecchio and Blough, 2004. doi: 10.1021/es049912h

Modeling "CDOM"

• Single-exponential function often describes the absorption magnitude and spectral dependence well, with two parameters:

$$a_{CDOM}(\lambda) = a_{CDOM}(\lambda_0) e^{-S_{CDOM}(\lambda - \lambda_0)}$$

- Steeper spectral slope is often interpreted as whether cDOM that is
 - Lower molecular weight
 - More photobleached/degraded
 - More "fulvic" in character (as opposed to "humic")
 - Operational definition from soil chemistry; based on solubility as a function of pH
- Does S_{CDOM} depend on the wavelength range of measurement?



Fig. 2. Representative absorption spectra, plotted as the natural logarithm of the absorption coefficient (m^{-1}) vs. wavelength. These spectra were collected in August 1991 on the transect between the central Gulf of Mexico and Oyster Bay.

Figure, Green and Blough 1994, Limnol. Ocean., 39(8):1903-1916.

Modeling "CDOM"

Spectra do not always follow a smooth or exponential functional shape!



Figure 8.15, Mobley et al., *The Oceanic Optics Book*, 2022. Data courtesy of A. Lyoub.

Non-pigmented particle absorption aka "non-algal particles" (NAP)

- Absorption by operationally defined particles
- Excluding the contribution by phytoplankton pigments



Extract the filter in hot methanol (or sodium hypochlorite, bleach) and measure its absorption (next lecture)



Figure 5.16: Diagrammatic representation of the geometric pathlength of the filtered sample. $V_{\rm filt}$ is the filtered volume, $d_{\rm eff}$ is the diameter of the circular distribution of particles on the filter, and is used to compute the effective area, $area_{\rm eff}$. The volume filtered can be expressed as a cylinder of area $area_{\rm eff}$ and length, ℓ . The geometric pathlength of the sample is ℓ .

Figure, C. Roesler, in Mobley et al. 2022, The Oceanic Optics Book

Contributors to a_{NAP}

- All phytoplankton cellular components that weren't soluble in the methanol
- All non-phytoplankton cells and organisms
- All non-living particles of biological origin
- All non-living mineral particles



Mineral contributions to $a_{NAP}(\lambda)$

112.

- $\frac{a}{[c]} \frac{m^{-1}}{3/m^3} = \frac{m^2}{g}$ Mineral samples with high iron have higher $a(\lambda)$ ٠
- Loss of a_{NAP}(UV-blue) "shoulders" along ٠ freshwater-marine sample transect
- Spectral derivatives: quantify the shoulders, then ٠ relate to electronic transitions of a specific subfraction of iron oxides present in the particles





Phytoplankton pigment absorption: $a_{\phi}(\lambda)$

- Measured by difference:
- Sum of all solvent-extractable pigments in the cell
- Pigments are "packaged" bound in membranes inside chloroplasts
- Not all pigments are extractable in methanol (water-soluble phycobiliproteins)



Figure 5.2: Generic phytoplankton absorption spectrum for mixed algal composition (modified from Roesler et al. (1989)).









Diatom (Coscinodiscus centralis)

Figure 1: Phytopedia, https://www.eoas.ubc.ca/research/phytoplankton/diatoms/centric/coscinodiscus/c_centralis.html Figures 2-3: JTO Kirk, *Light and Photosynthesis in Aquatic Ecosystems*, Ch. 8. Cambridge University Press, 2011. Figure 4 (spectrum): Fig. 8.10, Mobley et al. 2022, *The Oceanic Optics Book*.



Schematic of thylakoid membrane





TABLE 4.1. Summary of signature pigments useful as markers of algal groups and processes in the sea (see also Table 2.3, Chapter 2; and Chapter 13)^{a,b}

	Pignsent	Algal group or process	Key references	
	A. Chlorophylls (see Table 2.3, Chapter 2)			
	Chl a	All photosynthetic microalgae (except prochlorophytes)	See Table 2.3, Chapter 2	
Jeffrey, UNESCO 1997	Divinyl chl a	Prochlorophytes	Goericke & Repeta (1992)	
Summary table of	Chl b	Green algae: chlorophytes, prasinophytes, euglenophytes	See Table 2.3, Chapter 2	
, chlorophyll markor	Divinyl chl b	Prochlorophytes	Goericke & Repeta (1992)	
	Chl c family	Chromophyte algae	Jeffrey (1989)	
pigments for different	Chl c,	Diatoms, some prymnesiophytes, some freshwater	Jeffrey (1976b); Stauber & Jeffrey (1988); Jeffrey (1989);	
algal groups		chrysophytes, raphidophytes	Andersen & Mulkey (1983)	
0 0 1	Chl c ₂	Most diatoms, dinoflagellates, prymnesiophytes,	Jeffrey et al. (1975); Stauber & Jeffrey (1988); Andersen &	
		raphidophytes, cryptophytes	Mulkey (1983)	
Also: carotenoids,	Chl c3 ^s	Some prymnesiophytes, one chrysophyte, several diatoms	Jeffrey & Wright (1987); Vesk & Jeffrey (1987); Jeffrey	
biliproteins, many		and dinoflagellates	(1989); Johnsen and Sakshaug (1993)	
degradation products	Chl CONTROL	One prasinophyte	Jeffrey (1989)	
degradation products:	Phytylated chl c-like	Some prymnesiophytes	Nelson & Wakeham (1989); Jeffrey & Wright (1994)	
	Mg3.8 DVP	Some prasinophytes	Ricketts (1966); Jeffrey (1989)	
	Bacteriochlorophylls	Anoxic sediments	Repeta et al. (1989); Repeta and Simpson (1991)	

Class context: Week 1 roadmap

