





**Depth**  
Replicate

**COMMENTS**

S###

Z00 R01

m

Z00 R02

m

Z02 R01

m

Z02 R02

m

**Depth**

**COMMENTS**

P###

Z00

m

Z02

m

**Depth**

**COMMENTS**

S###L

Z00

m

Z02

m





Depth		COMMENTS TOC	COMMENTS DICTA	COMMENTS SAL
Z00	m	TOP FROM NISKIN ORDER: NUT-1,2,3; DOC1,2,3; DOM-1,2,3 <u>L -&gt; STERILE FILTER</u>	AT THE END THE FILTER WAS COLORED A LOT ALMOST CLOGGED ON <u>DOM-3</u>	
Z02	m			
Depth	Replicate	COMMENTS CDOM/FDOM	COMMENTS DOC	COMMENTS NUT
Z00	R01			
	m			
Z00	R02			
	m			
Z00	R03			
	m			
Z02	R01			
	m			
Z02	R02			
	m			
Z02	R03			
	m			





Depth		COMMENTS
Z00	R01 m	MB FILTRATION STARTED WITH 30 min DELAY (eDNA Protocol) ENGINES ON DURING THE EXTRACTION
Z00	R02 m	
Z00	R03 m	
Z00	R04 m	
Z02	R01 m	
Z02	R02 m	
Z02	R03 m	
Z02	R04 m	



## Tips for operator C

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- Margaux Crédaville (Ostende - Aarhus)

### Day before the station

- Prepare the logsheets, labels, tubes (only logsheets "samples" <sup>800</sup>  
print the logsheets "event" <sup>200</sup>)
- 2 racks with location of tubes are ready to use for 4 stations, in the wetlab <sup>m</sup>
- Few tubes are available in a grey box in the wetlab, near the peristaltic pump. If not enough, the stock is in the forepeak. <sup>200</sup>  
<sup>R07</sup>
- Make sure you have all the consumables you need for the station (enough Dacron pads and filters, enough bleach 10% for rinsing, spray bottles filled) <sup>m</sup> <sup>200</sup>  
<sup>R06</sup>

### Starting the station

- In the atelier, if not yet done switch ON the circuit-breaker of the wetlab on electric board <sup>200</sup>  
<sup>R05</sup>
- In the wetlab, prepare all the tripods
  - o Humidify the blue filter holders using spray of MQW and place Dacron pads <sup>200</sup>  
<sup>R04</sup>
  - o Humidify the Dacron pads using spray of MQW and place the right filters <sup>m</sup> <sup>200</sup>  
<sup>R05</sup>
  - o Close all the tripods <sup>200</sup>  
<sup>R03</sup>
- Outside the wetlab,
  - o Place the funnel equipped with 20 µm mesh on the dedicated holder. Attach using elastic a piece of 200 µm mesh on the outlet of the tube connected to pump A20. Then attach the weight on the inlet of the tube. Dip the tube inlet to the surface sea water (~1m) <sup>m</sup> <sup>200</sup>  
<sup>R01</sup>
  - o Start the pump A20 (switch O/I in the wetlab) and rinse all carboys using the FSW<20 µm. For virus carboys // and // rinse with FSW<0,1 µm. The pipe of the Filter Sea Water <sup>REPPLICATE</sup> <sup>COMMISSION</sup> <sup>200</sup> µm is on the right of the wetlab and the switch is located on the aft deck, port side <sup>Depth</sup> <sup>200</sup>  
<sup>R02</sup>





YYYY MM DD # # #  
 LOG\_SAMPLES\_ 2023 06 24 \_STATION\_ 0 4 1 \_S-LAB-DECKNET-5

OPERATOR(S) Agata .

Depth	DEPTH Volume (Litres)	Time start FILLING ##:##	Time end NET OUT ##:##	SG5-1* Cryo-5mL LN2	SG5-2* Cryo-5mL LN2
Z00 m	[ ] 100 L 60 L	04 : 32	09 : 00	112497108	112497109
Z02 m	[ ] 100 L L			### Z02 SG5-1	### Z02 SG5-2
Depth	FM5-1* Falcon-50mL FRG +4°C	FM5-2* Falcon-50mL FRG +4°C			
Z00 m	112557740	112557741			
Z02 m	### Z02 FM5-1	### Z02 FM5-2			
*pre-aliquoted 5 mL PFA/GLUT store at -20°C					
* pre-aliquoted Glycine betaine store at 4°C					

- When the first carboy is full, go to the next one and start the filtration immediately (Cf Handbook), launch the timer and the debimeter.
- Process in the same way for the next full carboy

### During the station

- Keep always an eye on carboys and tripods
- If air is present don't hesitate to use the bleeding valves
- If there are leaks around or under the tripod, stop the filtration by releasing the hose from the head and resolve it. The reasons can be various. Incorrectly tightened throttles, jaws or clamps. Damaged O-ring, filter support and/or grille badly positioned. After some time of filtration it can be due to the saturation of the filter. In this case you may have to decide to stop the filtration before reaching the set time or volume.  
All stuff and spare for reparation are in the black and red case in the wetlab
- Fill the logsheet with filtered volume, time of filtration and any comments (at the back of the logsheet) if something have running wrong

200 R02 m

### Ending the station

- Store S320-L/S023-L and S<02-R01 and R02 in the forepeak freezer and fridge respectively
- Clean all the tripods and carboys and let dried (tripode opened)
- Fill the MQW tank (it's not recommended to do that at the begining or during the station)
- Prepare new bleach solution (10%) for the next station
- Check your logsheets (Events and Samples) and store them in the folder « Logsheets to scan »
- Fill the TaraEuropa\_Samples\_Inventory\_Shipping file  
[https://docs.google.com/spreadsheets/d/180Bqgv3TUK45k79oEUiC7QQH7Rejp2MMH\\_K1KHyq\\_nc/edit#gid=0](https://docs.google.com/spreadsheets/d/180Bqgv3TUK45k79oEUiC7QQH7Rejp2MMH_K1KHyq_nc/edit#gid=0)

200 R02 m

SGS COMMENTS

Depth







Depth Replicate	COMMENTS PM	COMMENTS FOI
Z00 R01  m		
Z00 R02  m		
Z00 R03  m		
Z02 R01  m		
Z02 R02  m		
Z02 R03  m		

Depth Replicate	COMMENTS PA - HP
Z00  m	
Z02  m	





SAMPLE SPLITTING	COMMENTS	COMMENTS
PROTOCOLS		
S20 Cryo-5mL LN2 #1		
FCAM20 Bottle-250mL LIVE		
E20 Falcon-15mL FRZ -20°C		
S20-L Falcon-5mL FRZ -20°C		
MB20 Vial-4mL FRZ -20°C		
FM20 Falcon-50mL FRG +4°C		





	COMMENTS	COMMENTS
SAMPLE SPLITTING		
PROTOCOLS		
F200 Bottle-250mL RT >10°C		
SAMPLE SPLITTING		
PROTOCOLS		
S200 Cryo-5mL LN2 #1		
S200-L Falcon-5mL FRZ -20°C		



YYYY MM DD # # #  
 LOG-SAMPLES\_ 2023 06 24 \_STATION\_ 0 4 1 \_S-LAB-NET-680

OPERATOR(S) D. D.

Régent 680

SAMPLE SPLITTING	NET TOW #1			NET TOW #2		
	Total volume [ ] 1600 mL	Fraction of total volume	Bottle volume (mL)	Total volume [ ] 1600 mL	Fraction of total volume	Aliquot Volume (mL)
PROTOCOLS	Barcode			Barcode		
F680 Bottle-250mL RT >10°C + Borax/Formol	 112557671	[ ] 50 % [ ] 100 %	250 mL			
F2000 Bottle-250mL RT >10°C + borax/formol	### EPI F2000	hand-picked #ind=	[ ] 250 mL			
S680-L Falcon-5mL FRZ -20°C + 5mL Nucleoprotect				 112557672	[ ] 50 % [ ] 100 %	[ ] 200 mL [ ] 400 mL [ ] 600 mL [ ] 800 mL  250 mL [ ] 15 mn



	COMMENTS	COMMENTS
SAMPLE SPLITTING		
PROTOCOLS		
F680 Bottle-250mL RT >10°C		
F2000 Bottle-250mL RT >10°C		
S680-L Falcon-5mL FRZ -20°C		